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(54) Title: SKIN CARE COMPOSITIONS CONTAINING COMBINATIONS OF COMPOUNDS FOR MIMICKING THE EFFECT ON SKIN OF RETINOIC ACID (57) Abstract Compounds which sufficiently inhibit LRAT or ARAT catalyzed esterification of retinol into inactive retinyl esters, have the same effect on keratinocytes as retinoic acid. Thus, effects of retinol or retinyl esters in combination with these compounds are analogous to treatment with retinoic acid.		

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SKIN CARE COMPOSITIONS CONTAINING COMBINATIONS OF COMPOUNDS
FOR MIMICKING THE EFFECT ON SKIN OF RETINOIC ACID

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Field of the Invention

The present invention relates to skin care compositions containing certain compounds in combination with retinol and/or retinyl ester and to cosmetic methods involving applying such compositions to the skin.

Background of the Invention

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Retinol (vitamin A) is an endogenous compound which occurs naturally in the human body and is essential for normal epithelial cell differentiation. Natural and synthetic vitamin A derivatives have been used extensively in the treatment of a variety of skin disorders and have been used as skin repair or renewal agents. Retinoic acid has been employed to treat a variety of skin conditions, e.g., acne, wrinkles, psoriasis, age spots and discoloration. See e.g., Vahlquist, A. et al., *J. Invest. Dermatol.*, Vol. 94, Holland D.B. and Cunliffe, W.J. (1990), pp. 496-498; Ellis, C.N. et al., "Pharmacology of Retinols in Skin", *Vasel, Karger*, Vol. 3, (1989), pp. 249-252; Lowe, N.J. et al., "Pharmacology of Retinols in Skin", Vol. 3, (1989), pp. 240-248; PCT Patent Application No. WO 93/19743.

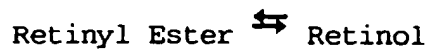
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It is believed that the use of retinol or esters of retinol would be preferred over retinoic acid. Retinol occurs naturally in the human body and is considered much safer than retinoic acid. Esters of retinol hydrolyze in-vivo to produce retinol. It is believed that retinol esters and retinol are

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metabolically converted in the skin into retinoic acid according to the following mechanism:



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Retinoic Acid

- 10 However, most of the endogenously applied retinol is rapidly converted into inactive fatty esters for storage in epidermal cells (keratinocytes). Esterification of retinol into inactive retinyl esters is achieved in cells by transfer of a fatty acyl group from an acyl CoA, catalyzed by the enzyme
- 15 acyl CoA retinol transferase (ARAT), or by the transfer of an acyl group from phosphatidyl choline, catalyzed by the enzyme lecithin retinol acyl transferase (LRAT). These esterification reactions are very efficient in keratinocytes--the majority (95%) of cellular retinoids are in the form of
- 20 retinyl fatty esters. Thus, unfortunately, although retinol and retinyl esters are safer to use than retinoic acid, they are less effective than retinoic acid at providing skin benefits.
- 25 The present invention is based, in part, on the discovery that certain compounds inhibit these esterification reactions and thus potentiate the action of retinol by increasing the amount of retinol available for conversion to retinoic acid. Thus, a mixture of these compounds with retinol or retinyl esters
- 30 mimics retinoic acid yet is safer to use than retinoic acid.

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Summary of the Invention

The present invention includes, in part, a skin conditioning composition containing:

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(a) from 0.001% to 10%, by weight of the composition of retinol or retinyl ester;

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(b) from 0.0001% to 50%, by weight of the composition of a compound which at 100 μ M concentration inhibits at least 20% of LRAT or ARAT catalyzed retinol esterification as measured by an in vitro Microsomal Assay; wherein the compound is not a fatty acid amide or dimethyl imidazolidinone; and

15

(c) a cosmetically acceptable vehicle.

Preferred compounds for use in the present invention are cyclic aliphatic unsaturated hydrocarbons, diterpenes, flavonones, flavonols, fatty hydroxyethyl imidazoline, surfactants and mixtures thereof. The compounds included in the present invention in combination with retinol and/or retinyl ester are selected based on the ability of such combinations to mimic retinoic acid's effect on skin. If the compound passes an in-vitro Microsomal Assay as described herein, it is included in the present invention even if it is not explicitly mentioned herein. Put another way, if a compound inhibits sufficiently an LRAT or ARAT catalyzed retinol esterification as measured by an in-vitro Microsomal Assay, it will act in combination with retinol or retinyl ester to mimic the effect on keratinocytes (skin cells) of retinoic acid.

The invention further provides a cosmetic method of conditioning skin comprising topically applying the present

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composition to the skin. The invention also provides a cosmetic method mimicking the effect of retinoic acid on skin, comprising topically applying the present composition to the skin.

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The term "conditioning" as used herein means prevention and treatment of one or more of the following: dry skin, photodamaged skin, appearance of wrinkles, age spots, aged skin. The compositions are also useful for increasing stratum corneum flexibility, lightening skin color, controlling sebum excretion and generally increasing the quality of skin. The composition may be used to improve skin desquamation and epidermal differentiation.

15 The presence of the selected compounds in the inventive composition substantially improves the performance of retinol or retinyl ester.

According to the present invention, by virtue of including an effective amount of a compound which at 100 μ M concentration inhibits at least 20% of ARAT or LRAT catalyzed retinol esterification as measured by in vitro Microsomal Assay, into compositions containing retinol or a retinyl ester, the performance of the compositions is substantially improved.

25

Description of the Preferred Embodiment

The inventive compositions contain, as a first essential ingredient, a compound selected from the group consisting of retinol and retinyl ester. The term "retinol" includes amongst others the following isomers of retinol: all-trans-retinol, 13-cis-retinol, 11-cis-retinol, 9-cis-retinol, 3,4-didehydro-retinol. Preferred isomers are all-trans-retinol, 13-cis-retinol, 3,4-didehydro-retinol, 9-cis-retinol. Most

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preferred is all-trans-retinol, due to its wide commercial availability.

Retinyl ester is an ester of retinol. The term "retinol" has
5 been defined above. Retinyl esters suitable for use in the
present invention are C₁-C₂₀ esters of retinol, preferably C₂-C₂₀
esters, and most preferably C₂, C₃, and C₁₆ esters because they
are more commonly available. Examples of retinyl esters
include but are not limited to: retinyl palmitate, retinyl
10 formate, retinyl acetate, retinyl propionate, retinyl
butyrate, retinyl valerate, retinyl isovalerate, retinyl
hexanoate, retinyl heptanoate, retinyl octanoate, retinyl
nonanoate, retinyl decanoate, retinyl undecanoate, retinyl
laurate, retinyl tridecanoate, retinyl myristate, retinyl
15 pentadecanoate, retinyl heptadecanoate, retinyl stearate,
retinyl isostearate, retinyl nonadecanoate, retinyl
arachidonate, retinyl behenate, retinyl linoleate, retinyl
oleate.

20 The preferred ester for use in the present invention is
selected from retinyl palmitate, retinyl acetate and retinyl
propionate, because these are the most commercially available
and therefore the cheapest. Retinyl linoleate is also
preferred due to its efficacy.

25 Retinol and/or retinyl ester is employed in the inventive
composition in an amount of from 0.001% to 10%, preferably in
an amount of from 0.01% to 1%, most preferably in an amount of
from 0.01% to 0.5%.

30 The second essential ingredient of the inventive compositions
is a compound which passes an in vitro Microsomal Assay. A
compound suitable for use in the present invention inhibits at
100 µM concentration, at least 20% of LRAT or ARAT catalyzed
35 retinol esterification as measured by in vitro Microsomal

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Assay. The in vitro Microsomal Assay employed for determining the suitability of the inclusion of the compound in the inventive compositions is as follows:

- 5 In a preferred embodiment of the invention, a compound is selected which, at a 100 μ M concentration, inhibits at least 40% and most preferably at least 50%, of LRAT or ARAT catalyzed retinol esterification.

10

In vitro Microsomal Assay:

- Microsomes are obtained as described in: J.C. Saari and D.L. Bredberg, "CoA and Non-CoA Dependent Retinol Esterification in
15 Retinal Pigment Epithelium" J. Biol. Chem. 263, 8084-90 (1988).

- A solution containing 0.1M sodium phosphate pH 7 buffer, 5mM dithiothreitol, 2 mg/ml bovine serum albumin, 40 micromolar
20 palmitoyl CoA, 40 micromolar dilauroyl phosphatidyl choline, 10 micromolar retinol and a test compound or a solvent blank, is incubated for 1 hour at 37°C with a microsomal fraction isolated from bovine retinal pigment epithelial cells. After incubation, the reaction is quenched by addition of an equal
25 volume of ethanol, and the retinyl esters formed (retinyl laurate from the LRAT catalyzed reaction and retinyl palmitate from ARAT catalyzed reaction) are extracted with hexane. The hexane layer is removed, evaporated under nitrogen, and the residue analyzed by HPLC on a 3.9x300 mm C₁₈ reversed phase
30 column using a 80% methanol in tetrahydrofuran mobile phase and fluorescence detection (325 nm excitation, 480 nm emission) to quantitate the retinyl ester. The quantity of ester formed in the presence of the solvent blank is taken as 100%, and this is used to calculate the percent inhibition of
35 ester formation for the compounds tested. As a control, an

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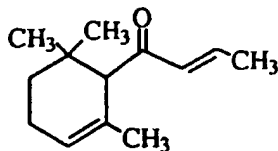
aliquot of microsomes is inactivated by boiling for 5 minutes, which results in at least 95% inhibition of ester formation.

- Cyclic aliphatic unsaturated hydrocarbons, diterpenes, flavonones, flavonols and certain fatty hydroxyethyl imidazoline surfactants are examples of the compounds which satisfy the in-vitro Microsomal Assay test described above. These compounds are described individually hereinbelow. Of course, other compounds not explicitly mentioned herein are included in the inventive compositions, as long as they pass the in-vitro Microsomal Assay Test described herein.

Cyclic Aliphatic Unsaturated Compounds

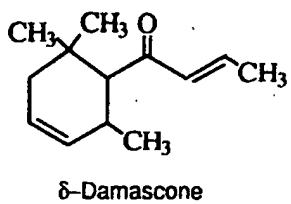
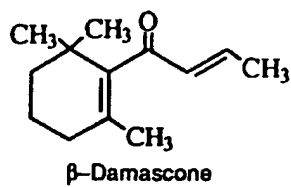
- Suitable cyclic aliphatic unsaturated compounds are selected according to the in-vitro Microsomal Assay Test described above.
- A preferred cyclic aliphatic unsaturated compound is selected from cyclic aliphatic unsaturated aldehydes, ketones, alcohols and esters.

- Preferred cyclic aliphatic unsaturated aldehydes, ketones, alcohols and esters are: alpha damascone, beta damascone, delta damascone, isodamascone, damascenone, alpha ionone, beta ionone, allyl alpha ionone, isobutyl ionone, alpha methyl ionone, gamma methyl ionone, brahmanol, sandanol, alpha terpineol, lylal, ethyl saffranate, and mixtures thereof.
- Structures of these compounds are as follows:

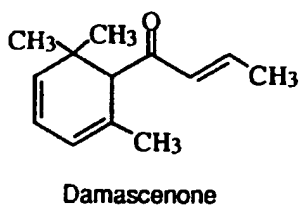
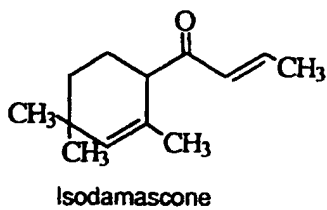


α -Damascone

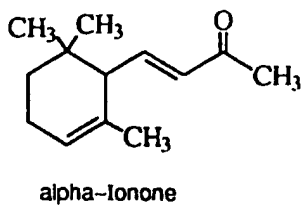
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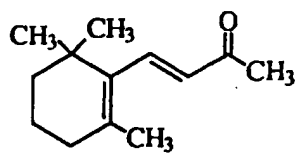
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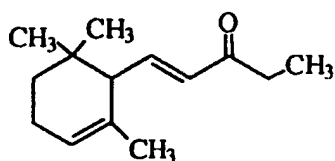
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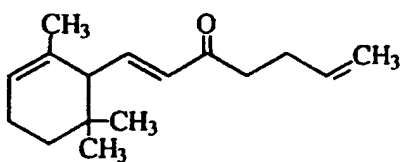


beta-Ionone



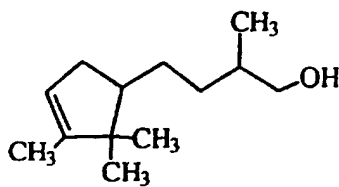
alpha-Methyl Ionone

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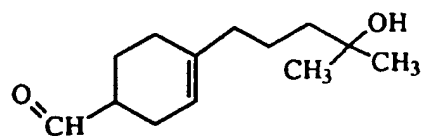
Allyl alpha-Ionone

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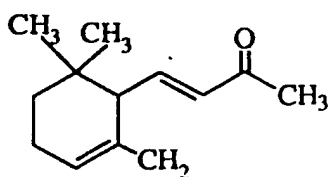


Brahmanol

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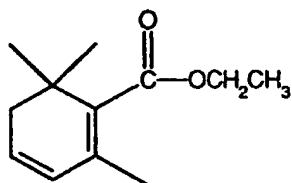


Lyrall



gamma-Methyl Ionone

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Ethyl Saffranate

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Preferably, in order to maximize performance at a minimum cost, a cyclic aliphatic unsaturated compound is selected from the group consisting of damascones and ionones having the structures described above.

Most preferably, the cyclic aliphatic unsaturated compound is a α -Damascone and/or α -Ionone.

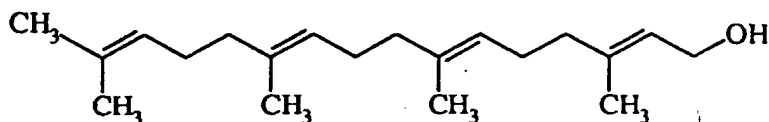
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The cyclic aliphatic unsaturated compound can be included in the inventive compositions in an amount ranging from 0.0001% to 50% by weight of the composition, preferably it is used in an amount of from 0.01% to 10%, most preferably from 0.1% to 5%.

Diterpenes

Diterpenes suitable for inclusion in the present invention are those that pass the in-vitro Microsomal Assay described hereinabove. A preferred diterpene compound is geranyl geraniol, which has the following structure:



Diterpene can be included in the inventive compositions in an amount ranging from 0.0001% to 50%, preferably it is used in an amount of from 0.01% to 10%, most preferably from 0.1% to 5%.

Flavonones and Flavonols

Flavonones and flavonols included in the present invention are those that pass in-vitro Microsomal Assay as described above.

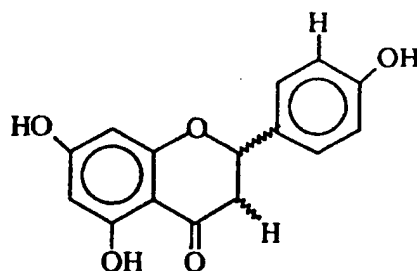
The preferred flavonones and flavonols are selected from naringenin, quercetin, and mixtures thereof.

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The structures of naringenin and quercetin are as follows:

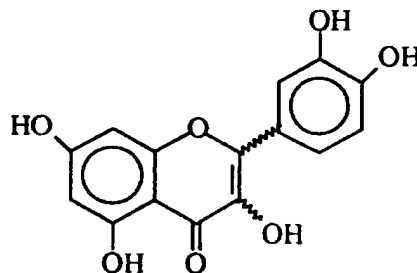
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Naringenin



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Quercetin

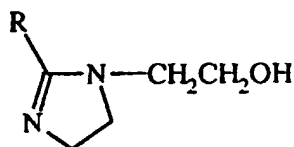


- 15 The flavonone or flavonol can be included in the inventive compositions in an amount ranging from 0.0001% to 50%, preferably it is used in an amount of from 0.01% to 10%, most preferably from 0.1% to 5%.
- 20 Quercetin and/or naringenin may be obtained from Sigma. Plant extracts containing quercetin and/or naringenin are also suitable for use in the present invention, e.g. rutin, evening primrose, onion, citrus species.

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Fatty Hydroxyethyl Imidazoline Surfactants

Fatty hydroxyethyl imidazoline surfactants included in the present invention pass the in-vitro Microsomal Assay test described above. Preferred fatty hydroxyethyl imidazolines have the following general structure:



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wherein R is an aliphatic saturated or unsaturated, straight or branched hydro-carbon chain containing from 8 to 20 carbon atoms.

15 Examples of suitable fatty hydroxyethyl imidazolines include but are not limited to cocoylhydroxyethyl imidazoline (R is derived from coconut oil fatty acids, mostly C₁₂ and some longer chain), lauryl hydroxyethyl imidazoline (R=CH₃(CH₂)₁₀), myristyl hydroxyethyl imidazoline (R=CH₃(CH₂)₁₂), stearyl
20 hydroxyethyl imidazoline (R=CH₃(CH₂)₁₆), and oleyl hydroxyethyl imidazoline (R=CH₃(CH₂)₇CH=CH(CH₂)₇).

Preferably, R in the fatty hydroxyethyl imidazoline contains from 8 to 18 carbon atoms, more preferably from 11 to 18
25 carbon atoms. Most preferably, the fatty hydroxyethyl imidazoline is oleyl hydroxyethyl imidazoline, due to its commercial availability and efficacy.

The fatty hydroxyethyl imidazoline can be included in the
30 inventive compositions in an amount ranging from 0.0001% to 50%, preferably it is used in an amount of from 0.01% to 10%, most preferably from 0.1% to 5%.

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Cosmetically Acceptable Vehicle

The composition according to the invention also comprises a cosmetically acceptable vehicle to act as a dilutant,
5 dispersant or carrier for the active ingredients in the composition, so as to facilitate their distribution when the composition is applied to the skin.

Vehicles other than or in addition to water can include liquid
10 or solid emollients, solvents, humectants, thickeners and powders. An especially preferred nonaqueous carrier is a polydimethyl siloxane and/or a polydimethyl phenyl siloxane. Silicones of this invention may be those with viscosities ranging anywhere from 10 to 10,000,000mm²/s (centistokes) at
15 25°C. Especially desirable are mixtures of low and high viscosity silicones. These silicones are available from the General Electric Company under trademarks Vicasil, SE and SF and from the Dow Corning Company under the 200 and 550 Series. Amounts of silicone which can be utilized in the compositions
20 of this invention range anywhere from 5% to 95%, preferably from 25% to 90% by weight of the composition.

The cosmetically acceptable vehicle will usually form from 5% to 99.9%, preferably from 25% to 80% by weight of the
25 composition, and can, in the absence of other cosmetic adjuncts, form the balance of the composition. Preferably, the vehicle is at least 50 wt.%, more preferably at least 80 wt.% water, by weight of the vehicle. Preferably, water comprises at least 50 wt.% of the inventive composition, most
30 preferably from 60 to 80 wt.%, by weight of the composition.

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Optional Skin Benefit Materials and Cosmetic Adjuncts

An oil or oily material may be present, together with an emulsifier to provide either a water-in-oil emulsion or an oil-in-water emulsion, depending largely on the average hydrophilic-lipophilic balance (HLB) of the emulsifier employed.

The inventive compositions preferably include sunscreens. Sunscreens include those materials commonly employed to block ultraviolet light. Illustrative compounds are the derivatives of PABA, cinnamate and salicylate. For example, octyl methoxycinnamate and 2-hydroxy-4-methoxy benzophenone (also known as oxybenzone) can be used. Octyl methoxycinnamate and 2-hydroxy-4-methoxy benzophenone are commercially available under the trademarks, Parsol MCX and Benzophenone-3, respectively. The exact amount of sunscreen employed in the emulsions can vary depending upon the degree of protection desired from the sun's UV radiation.

Another preferred optional ingredient is selected from essential fatty acids (EFAs), i.e., those fatty acids which are essential for the plasma membrane formation of all cells, in keratinocytes EFA deficiency makes cells hyperproliferative. Supplementation of EFA corrects this. EFAs also enhance lipid biosynthesis of epidermis and provide lipids for the barrier formation of the epidermis. The essential fatty acids are preferably chosen from linoleic acid, γ -linolenic acid, homo- γ -linolenic acid, columbinic acid, eicosa-(n-6,9,13)-trienoic acid, arachidonic acid, α -linolenic acid, timnodonic acid, hexaenoic acid and mixtures thereof.

Emollients are often incorporated into cosmetic compositions of the present invention. Levels of such emollients may range from 0.5% to 50%, preferably between 5% and 30% by weight of

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the total composition. Emollients may be classified under such general chemical categories as esters, fatty acids and alcohols, polyols and hydrocarbons.

- 5 Esters may be mono- or di-esters. Acceptable examples of fatty di-esters include dibutyl adipate, diethyl sebacate, diisopropyl dimerate, and dioctyl succinate. Acceptable branched chain fatty esters include 2-ethyl-hexyl myristate, isopropyl stearate and isostearyl palmitate. Acceptable
- 10 tribasic acid esters include triisopropyl trilinoleate and trilauryl citrate. Acceptable straight chain fatty esters include lauryl palmitate, myristyl lactate, oleyl eurate and stearyl oleate. Preferred esters include coco-
- 15 caprylate/caprate (a blend of coco-caprylate and coco-caprate), propylene glycol myristyl ether acetate, diisopropyl adipate and cetyl octanoate.

- Suitable fatty alcohols and acids include those compounds having from 10 to 20 carbon atoms. Especially preferred are
- 20 such compounds such as cetyl, myristyl, palmitic and stearyl alcohols and acids.

- Among the polyols which may serve as emollients are linear and branched chain alkyl polyhydroxyl compounds. For example,
- 25 propylene glycol, sorbitol and glycerin are preferred. Also useful may be polymeric polyols such as poly-propylene glycol and polyethylene glycol. Butylene and propylene glycol are also especially preferred as penetration enhancers.

- 30 Exemplary hydrocarbons which may serve as emollients are those having hydrocarbon chains anywhere from 12 to 30 carbon atoms. Specific examples include mineral oil, petroleum jelly, squalene and isoparaffins.

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Another category of functional ingredients within the cosmetic compositions of the present invention are thickeners. A thickener will usually be present in amounts anywhere from 0.1 to 20% by weight, preferably from 0.5% to 10% by weight of the composition. Exemplary thickeners are cross-linked polyacrylate materials available under the trademark Carbopol from the B.F. Goodrich Company. Gums may be employed such as xanthan, carrageenan, gelatin, karaya, pectin and locust bean gum. Under certain circumstances the thickening function may be accomplished by a material also serving as a silicone or emollient. For instance, silicone gums with viscosity in excess of 10 centistokes and esters such as glycerol stearate have dual functionality.

Powders may be incorporated into the cosmetic composition of the invention. These powders include chalk, talc, kaolin, starch, smectite clays, chemically modified magnesium aluminum silicate, organically modified montmorillonite clay, hydrated aluminum silicate, fumed silica, aluminum starch octenyl succinate and mixtures thereof.

Other adjunct minor components may also be incorporated into the cosmetic compositions. These ingredients may include coloring agents, opacifiers and perfumes. Amounts of these other adjunct minor components may range anywhere from 0.001% up to 20% by weight of the composition.

Use of the Composition

The composition according to the invention is intended primarily as a product for topical application to human skin, especially as an agent for conditioning and smoothening the skin, and preventing or reducing the appearance of wrinkled or aged skin.

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In use, a small quantity of the composition, for example from 1 to 100ml, is applied to exposed areas of the skin, from a suitable container or applicator and, if necessary, it is then spread over and/or rubbed into the skin using the hand or
5 fingers or a suitable device.

Product Form and Packaging

10 The topical skin treatment composition of the invention can suitably be formulated as a lotion, a cream or a gel. The composition can be packaged in a suitable container to suit its viscosity and intended use by the consumer. For example, a lotion or cream can be packaged in a bottle or a roll-ball
15 applicator, or a propellant-driven aerosol device or a container fitted with a pump suitable for finger operation. When the composition is a cream, it can simply be stored in a non-deformable bottle or squeeze container, such as a tube or a lidded jar.

20 The composition may also be included in capsules such as those described in U.S. Patent 5,063,057.

The invention accordingly also provides a closed container
25 containing a cosmetically acceptable composition as herein defined.

The following specific examples further illustrate the
invention.

30

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MATERIALS AND METHODS

Cell Culture:

5 Human keratinocytes, isolated from neonatal foreskin by
trypsin treatment were grown in Dulbecco Modification Eagle
(DME) Hams F12 (1:1) medium/10% fetal calf serum in the
presence of irradiated 3T3 mouse fibroblasts for establishing
dividing keratinocyte colonies. Cells were grown under the
10 above condition until their second passage and kept frozen for
future use. Frozen second passage keratinocytes were thawed
and plated into the above medium and grown for five days
before they were switched to a serum-free MCDB 153-based
medium keratinocyte growth medium (KGM) from Clonetics
15 Corporation, San Diego, CA, containing 0.15 mM Ca, or
keratinocyte serum-free media (KSFM) from GIBCO containing
0.09 mM Ca). On day 7, when the cells were 80-90% confluent,
they were trypsinized and plated in the serum-free medium for
the various experiments.

20

Thymidine Assay

³H-Thymidine Incorporation and Keratinocyte Proliferation

25

The incorporation of ³H-thymidine by cultured keratinocytes was
used as an assay of keratinocyte proliferation. Thymidine is
one of four deoxynucleosides which are the monomeric units of
DNA, the universal library of genetic information in the
animal kingdom. Prior to cell division of a somatic cell such
30 as a keratinocyte, the complete genome of the cell undergoing
cell division is replicated. This involves large scale DNA
synthesis by the cell and enables both daughter cells to
receive identical copies of the genetic material. When
35 ³H-thymidine is included in the culture media of keratinocytes

- 20 -

which are synthesizing DNA in preparation for cell division then the labelled nucleoside is incorporated into the newly synthesized DNA. The extent of incorporation of ³H-thymidine into a population of cells is proportional to the rate of DNA synthesis by this population of cells and therefore an indication of their cellular proliferation.

Keratinocytes (that were cultured as described above) were plated in 24 well plates at a density of 20,000 cells per well in 1 ml media. After incubation for four days or until the cells were 60-70% confluent, the media was changed. Test compounds were added (in triplicate) to the wells 24 hours after the media change, and four hours later 1μCi ³H-Thymidine in 50 μl media was added per well. Cells were incubated for a further 24 hours. Media was removed from the cells, 10% ice cold trichloroacetic acid (TCA) added and plates were incubated on ice for 30 minutes. Cells were washed five times with 5% TCA and allowed to dissolve in 500 μl 0.1M NaOH for at least one hour (usually overnight). The preparations were neutralized with 0.1M HCl; 50 μl of the cell preparation was used to determine total protein content. Disintegrations per minute (DPM) from ³H labelling of DNA was determined by liquid scintillation counting of 900μl of the cell preparation. Thymidine incorporation results were expressed as DPM/μg protein.

Transglutaminase Assay

30 Transglutaminase Assay and Keratinocyte Differentiation

During the process of terminal differentiation in the epidermis, a 15nm thick layer of protein, known as the cornified envelope (CE) is formed on the inner surface of the cell periphery. The CE is composed of numerous distinct

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proteins which have been cross-linked together by the formation of N^ε-(γ-glutamyl) lysine isodipeptide bonds catalyzed by the action of at least two different transglutaminases (TGases) expressed in the epidermis.

- 5 Transglutaminase I (TGase I) is expressed in abundance in the differentiated layers of the epidermis, especially the granular layer, but is absent in the undifferentiated basal epidermis. Thus TGase I is a useful marker of epidermal keratinocyte differentiation with high TGase I levels
- 10 indicating a more differentiated state. An ELISA based TGase I assay, using a TGase I antibody, was used to assess the state of differentiation of the cultured keratinocytes in the examples that follow.

- 15 For Example 1, the following procedure was used:

- Keratinocytes (cultured as described above) were plated in 96 well plates at a density of 3,000 cells per well in 200 μl media. After incubation for four days the media was changed
- 20 to media containing test compounds (six replicates per test). The cells were cultured for a further 72 hours after which time the media was aspirated and the plates stored at -70°C. Plates were removed from the freezer, and the cells washed with PBS. 100 μl sterile water was added and the cells were
- 25 freeze fractured by freezing at -70°C then thawing. The cells were incubated for one hour at room temperature (R/T) with PBS/3% BSA (wash buffer, bovine serum albumin), then rinsed with a fresh aliquot of wash buffer. Cells were incubated with 50 μl of primary antibodies monoclonal anti-human
- 30 transglutaminase mouse antibody (IgG) obtained from Biomedical Industries diluted 1:2,000 in wash buffer for one hour, 37°C then rinsed two times with wash buffer. Cells were then incubated with 50 μl of secondary antibody (Fab fragment, peroxidase conjugated anti-mouse IgG obtaining from Amersham)

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diluted 1:4,000 in wash buffer for one hour at 37°C, then rinsed two times with wash buffer. Cells were incubated with substrate solution (4 mg o-phenylene diamine and 3.3 µl 30% H₂O₂ in 10ml 0.1M citrate buffer pH 5.0) for five minutes, R/T, in darkness (under aluminum foil). The reaction was stopped by the addition of 50 µl 4N H₂SO₄. The absorbance of samples was read at 492nm in the plate reader. Out of the six replicates, four were treated with both antibodies, two were treated only with the secondary antibody (i.e., to determine background binding of enzyme conjugated Ab). TGase levels were determined by subtracting background from the readings from each treatment and determining mean ± s.d. for the replicates exposed to both antibodies.

For other Examples in which TGase I level was measured, the following procedure was used:

Keratinocytes (cultured as described above) were plated in 96 well plates at a density of 3,000 cells per well in 200µl of cell culture media. After incubation for four days, the media was changed to media containing test compounds (six replicates per test). The cells were cultured for a further 72 hours after which time the media was aspirated and the plates stored at -70°C. After the plates were removed from the freezer, the cells were further freezed fractured by freezing and thawing and then washed 3x with PBS. The cells were incubated for one hour at room temperature (R/T) with TBS/5% BSA buffer. Cells were then incubated with 100µl of monoclonal anti-human transglutaminase (IgG) mouse antibody (primary antibody) obtained from Biomedical Technologies Inc. diluted 1:2000 in TBS/1% BSA buffer for two hours at 37°C, and then rinsed six times with wash buffer (TBS/1% BSA/0.05% Tween-20). Cells were next incubated with 100µl of Fab fragment, peroxidase conjugated anti-mouse IgG antibody (secondary antibody) from

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Amersham diluted 1:4,000 in wash buffer for two hours at 37°C and then rinsed three times with wash buffer and three times with PBS. Cells were incubated with substrate solution (4mg o-phenylene diamine and 3.3µl 30% H₂O₂ in 10mL 0.1M citrate buffer, pH 5.0) for five minutes at R/T and in darkness (under aluminum foil). The reaction was stopped by the addition of 50µl 4N H₂SO₄. The absorbance of samples was read at 492nm in the plate reader. Out of the six replicates, four were treated with both antibodies, two were treated only with the secondary antibody (i.e., to determine the background binding of the enzyme conjugated antibody). Transglutaminase I levels were determined by subtracted background from the readings from each treatment and determining the mean ± s.d. for the replicates exposed to both antibodies.

DNA Assay

The level of TGase I detected after treatment of the cells could be influenced by cell number, i.e., the greater the number of cells the greater the level of TGase-I detected. The level of Tgase I was normalized to DNA content of the cells in the same well thus eliminating variation due to differences in cell number. DNA quantitation is a particularly useful indicator of cell number, including keratinocyte cell number, because each cell has to all intents and purposes an identical genome and therefore an identical quantity of DNA. The total DNA content of a well of cells therefore is directly proportional to the cell number in that well. Quantitation of DNA was used to normalize the TGase data to cell number.

Keratinocytes were plated in 96 well plates at a density of 3,000 cells per well in 200µl media. After incubation for four days the media was changed for media containing test

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compounds (6 replicates per test). The cells were cultured for a further 72 hours after which time the media was aspirated and the plates stored for at least 1.5 hours at -70°C. Plates were removed from the freezer and thawed for 30 minutes. 100µl/well of Hoechst dye (1µg/ml final concentration) was added and this was incubated for 15 minutes, covered and then read in a fluorimeter (ex. 360nm and em. 460nm). The dye solution was removed and the wells were rinsed with PBS in preparation for the TGase assay.

10

EXAMPLE 1

Retinoic acid is more effective than retinol at altering keratinocyte differentiation state

15

The effect on Transglutaminase levels normalized to DNA content of the cells after addition of retinoic acid (RA) and retinol (ROH) was examined and the results are shown in Table 1.

20

- 25 -

TABLE 1

Treatment	mean TGase/ DNA x 10 ⁻⁴ ± s.d (% control)	p value vs Control	p value vs 2.5x 10 ⁻⁷ M ROH	p value vs 2.5x 10 ⁻⁶ M ROH	p value vs 2.5x 10 ⁻⁵ M ROH
Control	2.44 ± 0.24 (100%)	-	0.001	0.001	0.001
2.5x10 ⁻⁷ M RA	0.16 ± 0.11 (7%)	0.001	0.001	0.001	0.001
2.5x10 ⁻⁷ M ROH	1.14 ± 0.22 (47%)	0.001	-	0.001	0.001
2.5x10 ⁻⁶ M RA	1.34 ± 0.40 (55%)	0.001	0.2	0.001	0.001
2.5x10 ⁻⁶ M ROH	1.89 ± 0.30 (77%)	0.001	0.001	-	0.001
2.5x10 ⁻⁵ M RA	1.87 ± 0.49 (77%)	0.001	0.001	0.784	0.001
2.5x10 ⁻⁵ M ROH	2.70 ± 0.59 (>100%)	0.001	0.001	0.001	-

n = 3

5

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All concentrations of retinoic acid tested, i.e., 2.5×10^{-7} M, 2.5×10^{-8} M and 2.5×10^{-9} M decreased keratinocyte differentiation over the ethanol control and did so to a significantly greater extent than each of the corresponding 2.5×10^{-7} M, 2.5×10^{-8} M and 2.5×10^{-9} M retinol treatments. The decrease in transglutaminase level was dose dependent for both retinoic acid and retinol. This is consistent with retinoic acid having a greater inhibitory effect on epithelial differentiation than retinol.

10

EXAMPLE 2

Method of in vitro microsomal esterification of retinol:

15 Microsomes are obtained as described in: J.C. Saari and D.L. Bredberg, "CoA and Non-CoA Dependent Retinol Esterification in Retinal Pigment Epithelium" J. Biol. Chem. 23, 8084-90 (1988).

A solution containing 0.1M sodium phosphate pH 7 buffer, 5mM dithiothreitol, 2 mg/ml bovine serum albumin, 40 micromolar palmitoyl CoA, 40 micromolar dilauroyl phosphatidyl choline, 10 micromolar retinol and a test compound or solvent blank, was incubated for 1 hour at 37°C with a microsomal fraction isolated from bovine retinal pigment epithelial cells. After incubation, the reaction was quenched by addition of an equal volume of ethanol, and the retinyl esters formed (retinyl palmitate from the ARAT catalyzed reaction, and retinyl laurate from the LRAT catalyzed reaction) were extracted with hexane. The hexane layer was removed, evaporated under nitrogen, and the residue analyzed by HPLC on a 3.9x300 mm C18 reversed phase column using a 80% methanol in tetrahydrofuran mobile phase and fluorescence detection (325 nm excitation, 480 nm emission) to quantitate the retinyl esters. The quantity of ester formed in the presence of the solvent blank was taken as 100%, and this was used to calculate the percent

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inhibition of ester formation for the compounds tested. As a control, an aliquot of microsomes was inactivated by boiling for 5 minutes, which resulted in at least 95% inhibition of ester formation.

5

The results that were obtained are summarized in Table 2.

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TABLE 2

	COMPOUND	CONCENTRATION (μ M)	% INHIB. ARAT	% INHIB. LRAT
5	oleyl hydroxyethyl imidazoline	100	90	95
	oleyl hydroxyethyl imidazoline	10	14	28
	caprylic hydroxyethyl imidazoline	100	-	8
	diazolidinyl urea	100	0	0
	thiamine	100	0	0
10	caffeine	100	0	0
	adenine	100	0	0
	phenyl benzimidazole sulfonic acid	100	0	0
	uracil	100	0	0
15	tryptophan	100	0	0
	cocoglutamate	100	0	0
	dimethyl cocamine oxide	100	0	0
	disodium cocamphodiacetate	100	0	0
	oleamidopropyl betaine	100	0	0
20	oleamine oxide	100	0	0
	lauroyl sarcosine	100	20	0

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It can be seen that the hydroxyethyl imidazoline surfactant is a potent esterification inhibitor, while other surfactants and other heterocyclic compounds were essentially inactive.

- Caprylic hydroxyethyl imidazoline ($R=CH_2(CH_2)_6$) did not sufficiently inhibit LRAT.

EXAMPLE 3

- Example 2 was repeated with various hydroxyethyl imidazoline surfactants, having R groups as indicated in Table 3. The results that were obtained are summarized in Table 3.

TABLE 3

R GROUP	CONCENTRATION (MICROMOLAR)	% INHIB. ARAT	% INHIB. LRAT
Oleyl	100	90	90
Oleyl	10	15	12
Lauryl	100	53	47
Lauryl	10	0	0
Coco	100	68	65
Coco	10	0	0

- The results in Table 3 show relative activity oleyl>coco>lauryl. Coco is better probably because it is not purely C_{12} , but has some longer chain alkyl groups as well.

- The data in Table 2 on caprylic hydroxyethyl imidazoline showed only 8% inhibition at 100 micromolar. Taken together the data clearly shows relative activity oleyl>coco>lauryl>>caprylic.

- 30 -

EXAMPLE 4

Transglutaminase Assay was conducted using oleyl hydroxyethyl
imidazoline (OHI), retinoic acid (RA), retinol (ROH), and
5 retinyl palmitate (RP).

The results that were obtained are summarized in Table 4.

TABLE 4

10	<u>Exp#</u>	<u>Compound(s)</u>	<u>Concentration (Micromolar)</u>	<u>% of Control</u>
	1	RA	0.25	35.1
15	1	ROH	0.25	76.2
	1	OHI	1	80.7
	1	ROH+OHI	0.25+1	45.1
	2	RA	0.25	43.4
20	2	ROH	0.025	91.6
	2	OHI	1	78.2
	2	ROH+OHI	0.025+1	57.9
	3	RA	0.25	45.9
25	3	RP	0.25	91.6
	3	OHI	1	91.1
	3	RP+OHI	0.25+1	71.6

30 It can be seen from the results in Table 4 that retinoic acid
substantially decreased keratinocyte differentiation whereas
retinol, oleyl hydroxyethyl imidazoline, and retinyl palmitate
did not affect keratinocyte differentiation when used alone,
or did so to a much smaller degree than retinoic acid.

35 In experiments 1 and 2, when retinol was combined with oleyl
hydroxyethyl imidazoline, the result was a synergistic
decrease in keratinocyte differentiation, to levels
approaching the effect of retinoic acid.

40

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In experiment 3, when oleyl hydroxyethyl imidazoline was combined with retinyl palmitate, the result was not as dramatic as in experiments 1 and 2, but nevertheless a synergistic decrease was observed.

5

This example also confirms that a compound which passed the in-vitro Microsomal Assay described herein, actually had an effect on keratinocytes when combined with retinol at a level similar to the effect of retinoic acid.

10

EXAMPLE 5

The in vitro Microsomal Assay Test was run on the compounds listed in Tables 5A and 5B.

15

The compounds in Table 5A were tested at a 100 μ M concentration. The compounds in Table 5B were tested at a 10 μ M concentration.

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TABLE 5A

COMPOUND	% INHIBITION, ARAT	% INHIBITION, LRAT
alpha damascone	83	98
beta damascone	84	92
delta damascone	87	95
isodamascone	80	92
damascenone	70	79
alpha ionone	45	49
beta ionone	22	24
allyl alpha ionone	22	36
isobutyl ionone	8	45
alpha methyl ionone	67	77
gamma methyl ionone	21	38
brahmanol	70	75
sandalol	15	43
alpha terpineol	26	25
timberol	34	33
lyral	76	71
tonalid	50	33
ethyl saffranate	51	49
traseolide	41	21
sandalone	23	12

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TABLE 5B

COMPOUND	% INHIBITION, ARAT	% INHIBITION, LRAT
alpha damascone	67	87
beta damascone	45	52
delta damascone	58	64
damascenone	23	29
allyl alpha ionone	16	17

5

It can be seen from the results in Tables 5A and 5B that certain cyclic aliphatic unsaturated compounds are potent inhibitors of LRAT and ARAT catalyzed retinol esterification.

10

COMPARATIVE EXAMPLE 6

The in-vitro Microsomal Assay test was conducted with additional cyclic aliphatic unsaturated compounds. The results that were obtained are summarized in Table 6.

15

The compounds in Table 6 were tested at a 100 μ M concentration.

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TABLE 6

COMPOUND	% INHIBITION, ARAT	% INHIBITION, LRAT
dihydro alpha ionone	13	18
alpha ionol	0	0
beta ionol	0	0
cinnamaldehyde	0	0
vanillin	0	0
eucalyptol	0	0
menthol	0	0
thymol	0	0
carvone	0	0
camphor	0	0
mentone	0	0
fenchyl alcohol	12	4
isocyclogeraniol	18	16
dimethyl ionone	0	9
delta methyl ionone	0	10

- 5 It can be seen from the results in Table 6 that not all cyclic aliphatic unsaturated compounds inhibit or sufficiently inhibit LRAT and ARAT catalyzed retinol esterification.

10 EXAMPLE 7

The effect on keratinocyte differentiation of compounds and combinations listed in Table 7 was examined. The results were expressed as % of control. Transglutaminase level was
15 normalized to DNA. Data are from two experiments where the

- 35 -

retinol concentration was changed. The results that were obtained are summarized in Table 7.

5

TABLE 7

EXPERIMENT #	TREATMENT	CONCENTRATIONmM	% CONTROL
1	Retinoic acid	0.00025	24
1	Retinol	0.001	67
1	α -Damascone	1	92
1	Retinol + α -Damascone	0.001 + 1	34
2	Retinoic acid	0.00025	8
2	Retinol	0.00025	63.5
2	α -Damascone	1	86
2	Retinol + α -Damascone	0.00025 + 1	30

The results in Table 7 show that while α -Damascone alone and retinol alone were not very effective, the combination of the two attained synergistic reduction in transglutaminase mimicking the effect of retinoic acid on keratinocyte differentiation. This example also establishes a good correlation between the Microsomal Assay and cell culture data.

10

15

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EXAMPLE 8

The in-vitro Microsomal Assay test was conducted with a diterpene compound, geranyl geraniol or farnesol.

The results that were obtained are summarized in Table 8.

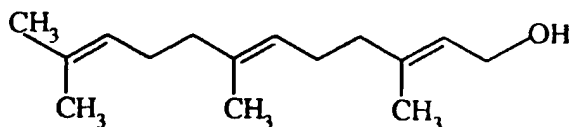
TABLE 8

COMPOUND	CONCENTRATION (μ M)	% INHIB. ARAT	% INHIB. LRAT
Geranyl Geraniol ¹	100	81	77
Geranyl Geraniol	10	38	16
Farnesol ²	100	43	43
Farnesol	10	20	10

¹ Obtained from TCI America (Portland, Oregon). Also available from Sigma and CTC Organics (Atlanta, Georgia).

² Available from Givaudan Co., Bedoukian Co., or Dragoco Co.

It can be seen from the results in Table 8 that both geranyl geraniol and farnesol inhibit retinol esterification. Geranyl geraniol is a substantially more potent esterification inhibitor, than farnesol, the structure for which is as follows:



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EXAMPLE 9

³H-Thymidine incorporation was measured according to the procedure described in Materials and Methods section above.

- 5 The results that were obtained are summarized in Tables 9A and 9B.

TABLE 9A

10

Experiment #1			
Treatment	DPM/Microgram Protein		% Control
	<u>mean</u>	<u>(s.d.)</u>	
Control		(278)	100
250 nM retinol	2082	(146)	108
250 nM retinoic acid	3013	(226)	156
250nM retinol + 10 nM geranyl geraniol	2970	(308)	154

TABLE 9B

15

Experiment #2			
Treatment	DPM/Microgram Protein		% Control
	<u>mean</u>	<u>(s.d.)</u>	
Control		(322)	100
250 nM retinol	974	(148)	107
250 nM retinoic acid	1494	(45)	163
250nM retinol + 10 nM geranyl geraniol	1450	(318)	159

It can be seen from the results above that geranyl geraniol is able to significantly increase the proliferation enhancing

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effect of retinol to a level similar to that which can be achieved with a comparable amount of retinoic acid. Again, a good correlation is proven to exist between the Microsomal Assay test and the effect of a compound on cell culture.

5

EXAMPLE 10

The in-vitro Microsomal Assay was conducted on compounds listed in Table 10. The results that were obtained are summarized in Table 10. The compounds in Table 10 were tested at 100 μ M concentration.

15

TABLE 10

COMPOUND	% INHIBITION, ARAT	% INHIBITION, LRAT
Control	0	0
Naringenin	33	14
Quercetin	25	14

EXAMPLE 11

20

Naringenin and Retinol Synergistically Inhibit Keratinocyte Differentiation

The effect on TGase I levels normalised to DNA content of the cells was examined in response to a 72 hour treatment with the test compounds. The results are shown in Table 11.

25

Naringenin was obtained from Sigma.

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TABLE 11

Effect of Retinol and Naringenin on Keratinocyte TGase/DNA

Treatment	mean TGase/ DNA x10 ³ ± s.d (% control)	p value vs Control	p value vs 2.5x 10 ⁻⁶ M ROH	p value vs 2.5x 10 ⁻⁶ M RA	p value vs 10 ⁻⁶ M naringenin
Control	52.78 ± 5.69 (100%)	-	0.235	0.001	0.329
2.5x10 ⁻⁷ M RA	22.47 ± 2.31 (42%)	0.001	0.001	-	0.001
2.5x10 ⁻⁹ M Retinol	48.31 ± 5.31 (92%)	0.235	-	0.001	0.585
10 ⁻⁷ M Naringenin	49.84 ± 2.76 (94%)	0.329	0.585	0.001	-
2.5x10 ⁻⁹ M ROH + 10 ⁻⁷ M Naringenin	27.61 ± 10.79 (53%)	0.002	0.005	0.328	0.002

n = 3

- 40 -

It can be seen from the results in Table 11 that 2.5×10^{-7} M retinoic acid was very effective at repressing keratinocyte TGase I levels (to 42% of control level). 2.5×10^{-5} M retinol was ineffective (91%) and 10^{-7} M naringenin had no inhibitory effect on the keratinocyte TGase I level when used alone. However, 2.5×10^{-5} M retinol + 10^{-7} M naringenin repressed keratinocyte TGase I to 53% of control levels. Naringenin and retinol therefore acted synergistically to repress keratinocyte differentiation in an analogous manner to the effect of retinoic acid.

EXAMPLE 12

15 Naringenin and Retinyl Palmitate Synergistically Inhibit Keratinocyte Differentiation

The effect on TGase I levels normalised to DNA content of the cells was examined in response to a 72 hour treatment with the test compounds. The results are shown in Table 12.

TABLE 12
Effect of Retinyl Palmitate and Naringenin on Keratinocyte TCase/DNA

Treatment	mean TCase/ DNA x10 ³ ± s.d (% control)	p value vs Control	p value vs 2.5x 10 ⁻⁶ M RP	p value vs 2.5x 10 ⁻⁷ M RA	p value vs 10 ⁻⁶ M naringenin
Control	50.64 ± 1.74 (100%)	-	0.143	0.001	0.001
2.5x10 ⁻⁷ M RA	16.31 ± 2.58 (32%)	0.001	0.001	-	0.001
2.5x10 ⁻⁹ M Retinyl Palmitate (RP)	47.32 ± 4.22 (93%)	0.143	-	0.001	0.296
10 ⁻⁶ M Naringenin	45.01 ± 1.90 (89%)	0.001	0.296	0.001	-
2.5x10 ⁻⁹ M RP + 10 ⁻⁶ M Naringenin	43.12 ± 13.01 (85%)	0.005	0.036	0.001	0.065

n = 3

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It can be seen from the results in Table 12 that 2.5×10^{-7} M retinoic acid was very effective at repressing keratinocyte TGase I levels (to 32% of control level). 2.5×10^{-3} M retinyl palmitate was ineffective (93%) and 10^{-8} M naringenin had a small inhibitory effect on the keratinocyte TGase I level when used alone. However, 2.5×10^{-8} M retinol + 10^{-8} M naringenin repressed keratinocyte TGase I to 85% of control levels. Naringenin and retinyl palmitate therefore act synergistically to repress keratinocyte differentiation in an analogous manner to the effect of retinoic acid.

EXAMPLE 13

15 Quercetin and Retinol Synergistically Inhibit Keratinocyte Differentiation

The effect on TGase I levels normalised to DNA content of the cells was examined in response to a 72 hour treatment with the test compounds. The results are shown in Table 13. Quercetin was obtained from Sigma.

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TABLE 13

Treatment	mean TGase/ DNA $\times 10^3 \pm \text{s.d.}$ (% control)	p value vs Control	p value vs 2.5×10^{-7} M ROH	p value vs 2.5×10^{-7} M RA	p value vs 10^{-6} M quercetin
Control	69.16 ± 4.26 (100%)	-	0.042	0.001	0.003
2.5×10^{-7} M RA	35.91 ± 3.01 (52%)	0.001	0.001	-	0.001
2.5×10^{-7} M Retinol	61.93 ± 5.18 (90%)	0.042	-	0.001	0.328
10^{-6} M Quercetin	59.04 ± 3.38 (85%)	0.003	0.328	0.001	-
2.5×10^{-7} M ROH + 10^{-6} M Quercetin	48.45 ± 8.60 (70%)	0.001	0.017	0.015	0.034

n = 3

5

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It can be seen from the results in Table 13 that $2.5 \times 10^{-7} \text{M}$ retinoic acid was effective at repressing keratinocyte TGase I levels (to 52% of control level). $2.5 \times 10^{-7} \text{M}$ retinol was ineffective (90%) and 10^{-6}M quercetin had only a small
5 inhibitory effect on the keratinocyte TGase I level when used alone. However, $2.5 \times 10^{-7} \text{M}$ retinol + 10^{-6}M quercetin repressed keratinocyte TGase I to 70% of control levels. Quercetin and retinol therefore acted synergistically to repress
10 keratinocyte differentiation in an analogous manner to the effect of retinoic acid.

During the course of these studies, retinoic acid was used as positive control and reference compound against which the
15 other compounds under analysis were compared. Retinoic acid, in a dose dependant manner decreased transglutaminase I levels in skin keratinocytes. In other words, retinoic acid decreased keratinocyte differentiation. Retinol and retinyl palmitate were significantly less effective than retinoic acid at inhibiting keratinocyte differentiation.

20 The unexpected result demonstrated by the Examples above however was that the effect of retinol or retinyl ester on cultured keratinocytes can be enhanced to levels approaching those of retinoic acid by combining retinol or the ester with
25 a compound which is from 0.0001% to 50% of a compound which at 100 μM concentration inhibits at least 20% of LRAT or ARAT catalyzed retinol esterification as measured by the in vitro Microsomal Assay. This effect was not only greater than the effect of either retinol or the ester or the compound itself
30 but the two ingredients acted in synergy with each other to promote a retinoic acid-type response on the keratinocytes.

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The results documented above demonstrate that compounds which pass the in-vitro Microsomal Assay act synergistically with retinol and retinyl esters to decrease keratinocyte differentiation and/or to increase keratinocyte proliferation, mimicking the effect on keratinocytes of retinoic acid.

Examples 14-19 illustrate topical compositions according to the present invention. The compositions can be processed in conventional manner. They are suitable for cosmetic use.

10 In particular the compositions are suitable for application to wrinkled, rough, dry, flaky, aged and/or UV-damaged skin to improve the appearance and the feel thereof as well as for application to healthy skin to prevent or retard deterioration thereof.

15

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EXAMPLE 14

This example illustrates high internal phase water-in-oil emulsions incorporating the inventive composition.

5

	% w/w			
	A	B	C	D
Retinol	0.5	---	---	0.01
Retinyl Palmitate	---	0.15	0.15	---
Fully hydrogenated coconut oil	3.9	3.9	3.9	3.9
Naringenin	---	5	0.1	5
Quercetin	1	1	---	---
Brij 92*	5	5	5	5
Bentone 38	0.5	0.5	0.5	0.5
MgSO ₄ ·7H ₂ O	0.3	0.3	0.3	0.3
Butylated hydroxy toluene	0.01	0.01	0.01	0.01
Perfume	qs	qs	qs	qs
Water	to 100	to 100	to 100	to 100

* Brij 92 is polyoxyethylene (2) oleyl ether

10

EXAMPLE 15

This example illustrates oil-in-water creams according to the invention.

5

	% w/w			
	A	B	C	D
Retinyl Palmitate	---	0.15	0.15	---
Retinol	0.15	---	0.001	0.15
Mineral oil	4	4	4	4
α -ionone	1	---	---	1
α -methylionone	---	0.5	---	---
Lyrar	----	---	---	0.2
Isodamascone	---	---	0.3	---
Brij 56*	4	4	4	4
Alfol 16RD*	4	4	4	4
Triethanolamine	0.75	0.75	0.75	0.75
Butane-1,3-diol	3	3	3	3
Xanthan gum	0.3	0.3	0.3	0.3
Perfume	qs	qs	qs	qs
Butylated hydroxy toluene	0.01	0.01	0.01	0.01
Water	to 100	to 100	to 100	to 100

* Brij 56 is cetyl alcohol POE (10)
Alfol 16RD is cetyl alcohol

10

EXAMPLE 16

This example illustrates alcoholic lotions according to the invention.

5

	% w/w			
	A	B	C	D
Retinol	---	0.15	0.15	---
Retinyl palmitate	0.15	---	----	0.15
α -Damascone	0.1	---	0.1	---
Geranyl Geraniol	---	1	---	0.2
Ethanol	40	40	40	40
Perfume	qs	qs	qs	qs
Butylated hydroxy toluene	0.01	0.01	0.01	0.01
Water	to 100	to 100	to 100	to 100

EXAMPLE 17

This example illustrates another alcoholic lotion containing the inventive composition.

10

	% w/w
Retinol	0.15
Oleyl Hydroxyethylimidazoline	0.1
Ethanol	40
Antioxidant	0.1
Perfume	qs
Water	to 100

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EXAMPLE 18

This example illustrates a suncare cream incorporating the composition of the invention:

5

	% w/w
Retinyl Palmitate	0.01
Cocoylhydroxyethylimidazoline	0.1
Silicone oil 200 cts	7.5
Glycerylmonostearate	3
Cetosteryl alcohol	1.6
Polyoxyethylene-(20)-cetyl alcohol	1.4
Xanthan gum	0.5
Parsol 1789	1.5
Octyl methoxycinnate (PARSOL MCX)	7
Perfume	qs
Color	qs
Water	to 100

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EXAMPLE 19

This example illustrates a non-aqueous skin care composition incorporating the inventive combination.

5

	% w/w
Retinyl Palmitate	0.15
Lauryl Hydroxyethyl imidazoline	1
Silicone gum SE-30 ¹	10
Silicone fluid 345 ²	20
Silicone fluid 344 ³	55.79
Squalene	10
Linoleic acid	0.01
Cholesterol	0.03
2-hydroxy-n-octanoic acid	0.7
Vitamin E linoleate	0.5
Herbal oil	0.5
Ethanol	2

10 ¹ A dimethyl silicone polymer having a molecular weight of at least 50,000 and a viscosity of at least 10,000 centistokes at 25°C, available from GEC

² Dimethyl siloxane cyclic pentamer, available from Dow Corning Corp.

15 ³ Dimethyl siloxane tetramer, available from Dow Corning Corp.

20 If not mentioned in the Examples, materials employed in the present invention are obtained from the following sources:

Palmitoyl CoA, BSA, dilauroylphosphatidyl choline, retinol, retinoic acid dithiothreitol - from Sigma.

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Cyclic aliphatic unsaturated compounds:

Damascones from Firmenich;

Ionones from IFF;

Others from suppliers in "Flavor and Fragrance

5 Materials" 1991, by Allured Pub. Co.

Imidazoline surfactants:

The oleyl imidazoline was Schercozoline O from Scher
Chemical Co.; The others were from McIntyre, under the10 Mackazoline name, as follows: Caprylic, Coco, Lauryl:
Mackazoline CY, C, and L, respectively.

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CLAIMS

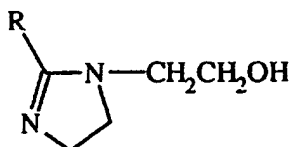
1. A skin care composition comprising:
 - 5 (a) from 0.001% to 10% of a compound selected from the group consisting of retinol, retinyl ester and mixtures thereof;
 - 10 (b) from 0.0001% to 50% of a compound which at 100 μ M concentration inhibits at least 20% of LRAT or ARAT catalyzed retinol esterification as measured by an in vitro Microsomal Assay; wherein the compound is not a fatty acid amide or dimethyl imidazolidinone; and
 - 15 (c) a cosmetically acceptable vehicle.
2. The composition of claim 1 wherein the compound (b) is
20 selected from the group consisting of cyclic aliphatic unsaturated hydrocarbons, diterpenes, flavonones, flavonols, fatty hydroxyethyl imidazoline surfactants and mixtures thereof.
3. The composition of claim 2 wherein the cyclic aliphatic
25 unsaturated compound is selected from the group consisting of alpha damascone, beta damascone, delta damascone, isodamascone, damascenone, alpha ionone, beta ionone, allyl alpha ionone, isobutyl ionone, alpha methyl ionone, gamma methyl ionone, brahmanol, sandanol, alpha terpineol, lyral, ethyl saffranate, and
30 mixtures thereof.
4. The composition of claim 2 or claim 3 wherein the
35 diterpene is geranyl geraniol.

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5. The composition of any one of claims 2-4 wherein the flavonone is naringenin.

6. The composition of any one of claims 2-5 wherein the flavonol is quercetin.

7. The composition of any one of claims 2-6 wherein the fatty hydroxyethyl imidazoline surfactant has the following structure:



wherein R is an aliphatic saturated or unsaturated, straight or branched hydro-carbon chain containing from 8 to 20 carbon atoms.

8. The composition of any one of claims 1-7 wherein the retinyl ester is selected from the group consisting of retinyl palmitate, retinyl acetate, retinyl propionate, retinyl linoleate and mixtures thereof.

9. A cosmetic method of conditioning skin the method comprising applying topically to skin a composition according to any one of claims 1-8.

10. A cosmetic method of mimicking the effect on skin of retinoic acid, the method comprising applying to the skin a composition according to any one of claims 1-8.

INTERNATIONAL SEARCH REPORT

International Application No

PCT/EP 97/05150

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 A61K7/48

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X,P	US 5 665 367 A (BURGER ET AL.) 9 September 1997 see the whole document	1,2,5,6, 8-10
X	WO 96 03973 A (LIFEGROUP SPA) 15 February 1996 see example 19	1,2,5,6, 8-10
X	FR 2 666 226 A (THOREL) 6 March 1992 see page 5, line 30 - line 39; examples 1-4	1,2,8-10
X	EP 0 601 698 A (SHISEIDO) 15 June 1994 see example 4	1,2,8-10
X	EP 0 608 433 A (SHISEIDO) 3 August 1994 see examples 2-3,2-8	1,2,8-10
	-/--	

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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"P" document published prior to the international filing date but later than the priority date claimed

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"&" document member of the same patent family

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INTERNATIONAL SEARCH REPORT

International Application No

PCT/EP 97/05150

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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A	<p>US 5 536 740 A (GRANGER ET AL.) 16 July 1996 see the whole document</p>	1,2,7-10
A	<p>FR 2 558 058 A (P.F.MEDICAMENT) 19 July 1985 see the whole document</p>	1,2,7-10

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Information on patent family members

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